

Executive Summary

Extrinsic atopic dermatitis, allergic asthma, and allergic rhinitis are prevalent allergic diseases. These diseases can be distinguished by the location of their most important symptoms; skin, lungs and sinuses, respectively. They also share many characteristics, including the same underlying immune system dysregulation as evidenced by inappropriate T-cell mediated responses to innocuous antigens, peripheral eosinophilia, elevated serum IgE levels, chronic immune system activation, and over-production of inflammatory mediators, including leukotrienes, cytokines and chemokines.

The role of inflammatory mediators in allergic diseases is well characterized. In particular, leukotrienes, derived from the fatty acid arachidonic acid, play a significant role in the pathophysiology of allergic diseases. There are two classes of leukotrienes with different roles. Cysteinyl leukotrienes (LTC_4 , LTD_4 and LTE_4) directly promote allergic inflammation in target tissues and increase the synthesis of other proinflammatory mediators, such as cytokines. The chemotactic leukotriene, LTB_4 , is a chemoattractant for immune cells and activates neutrophils leading to endothelial leakage.

It has been demonstrated that two different strategies to reduce the impact of leukotrienes on symptoms of allergic disease are effective: 1) blocking leukotriene receptors and 2) inhibiting the action of the leukotriene synthesis enzyme, 5-lipoxygenase (5-LO). The receptor blocking drugs target only one of the two classes of leukotrienes, yet appear to have a modest but clinically significant impact on disease symptoms. The leukotriene synthesis blocker blunts production of both classes of leukotrienes and also has a clinically significant impact on disease symptoms of atopic dermatitis (eczema), asthma and allergic rhinitis.

EFFICAS CARE is a specially formulated medical food based upon bioactive fatty acids with modes of action relevant for the dietary management of allergic diseases via reduction of leukotrienes. EFFICAS CARE has been shown in clinical trials to alter the pools of fatty acid precursors of leukotrienes, to significantly reduce the synthesis of leukotrienes by immune cells *ex vivo* in healthy individuals as well as in asthmatics. Efficas Care has also been shown during consumer trials to significantly improve disease management and the quality of life in individuals with asthma, allergic rhinitis and eczema. Since EFFICAS CARE blocks leukotriene synthesis and improves disease management, it is scientifically valid to target multiple allergic diseases with this product.

The Features of Allergic Disease.

Allergic disease is a dysregulation of the immune system, characterized by

1. Inappropriate T-cell responses
2. Tissue edema
3. Imbalances in the numbers, locations and activation status of specific immune cell types and subtypes
4. Allergen sensitivity and production of allergen specific IgE, and
5. Overproduction of and hypersensitivity to inflammatory mediators, including leukotrienes and cytokines

Extrinsic atopic dermatitis (AD), allergic asthma (AS), and allergic rhinitis (AR) are prevalent allergic diseases that can be distinguished by the location of their most important symptom manifestations; skin, lungs and sinuses, respectively. These diseases have the same underlying, systemic origin and two or more of them may occur simultaneously or sequentially in the same individual (Kagi, 2001; Leung et al. 2004; Meltzer, 2000; Oettgen and Geha, 1999; Spergel and Paller, 2003). Allergic disease normally begins in infancy, with either food allergy or AD. Approximately 50-80% of children with AD will go on to develop allergic rhinitis and/or asthma, a phenomenon known as the atopic march (Rackal and Vender, 2004; Spergel and Paller, 2003). The systemic origin is a dysregulation of the immune system response to environmental antigens, mediated by T-cells (Borish and Steinke, 2003). Allergic conditions are characterized by elevated serum IgE levels, peripheral eosinophilia, T-helper lymphocyte imbalances, markers of immune system activation such as mast cell activation and excess synthesis of leukotrienes and cytokines.

Imbalances in Immune cell types and subtypes

A brief discussion of immune cells involvement in allergic disease is given below. Consult the references for more detail.

T-Helper Cell Subtypes

In allergic disease, Th-2 cells predominate over Th-1 cells. This chronic state of T cell imbalance leads to the overproduction of inflammatory cytokines such as IL-4, IL-5 and IL-13. IL-4 in particular contributes to symptoms of itch, as well as stimulating IgE production by B cells. IL-5 stimulates production of eosinophils. T-cell derived cytokines also contribute to proliferation of mast cells.

Eosinophils

Excess numbers of activated eosinophils in target tissues and in circulation is a hallmark of all allergic diseases (Borish and Steinke, 2003; Prussin and Metcalfe, 2003). These cells are the major source of inflammatory cysteinyl leukotrienes and also produce IL-13. Leukotrienes cause bronchoconstriction in the lungs, mucous production in sinuses, and promote permeability to inflammatory cells in skin. IL-13 induces airway hyperresponsiveness (Vladich et al., 2005). Eosinophils also release major basic protein, which is toxic to epithelial cells in the respiratory tract.

Mast Cells

Allergic individuals produce greater numbers of mast cells than nonatopic individuals (Borish and Steinke, 2003; Prussin and Metcalfe, 2003). Mast cells bear allergen specific IgE. When exposed to an allergen, mast cells become activated and degranulate, releasing histamine, cytokines and prostaglandins and leukotrienes (Holgate, 2000). Mast cell degranulation quickly produces the typical symptoms of allergic disease: itch, redness and swelling of the skin; bronchoconstriction, mucous production, swelling and cough in the lungs; itch, sneezing and mucous production in sinuses.

Basophils

Basophils produce inflammatory mediators, including histamine, leukotriene LTC₄, IL-4, and IL-13 (Prussin and Metcalfe, 2003). Basophils, like mast cells, express high-affinity IgE receptor. When activated, basophils release histamine, cytokines and leukotrienes. Elevated levels of circulating basophils are characteristic of atopic asthma and atopic dermatitis. LTB₄ induces degranulation of IL-13 primed basophils.

Langerhan's cells.

Skin of individuals with active AD, AS and/or AR contains Langerhan's cells expressing surface FcεR1 with bound IgE (Semper et al., 2003). Thus, skin abnormalities exist in all allergic disease, regardless of whether AD is present.

The role of inflammatory mediators in allergic disease

Table 1 summarizes the involvement of key cytokines and leukotrienes in allergic disease

Table 1. Inflammatory mediators in allergic disease.

Cytokine	Source	Impacts in allergic disease
IL-4	Th-2 lymphocytes, eosinophils, basophils	Enhances antigen-presenting capacity of B cells. Induces isotype switching from IgM to IgE. Influences T-cell differentiation and survival. Induces mast cell expression of LTC ₄ synthase. Stimulates mucous production in airways. Stimulates production of leukotriene receptors.
IL-5	Th-2 lymphocytes	Promotes eosinophil differentiation and survival. Upregulates 5-LO
IL-13	Th-2 lymphocytes	Promotes IgE synthesis Promotes mucous secretion in airways.
Leukotriene		
LTB ₄	Neutrophils, macrophages	Chemoattractant for other immune cells in respiratory tract and skin, which prolongs the inflammatory response in these tissues. Activates neutrophils leading to endothelial leakage and promotion of inflammation. Promotes synthesis of IL-5 by T cells. Induces degranulation of basophils.
LTC ₄ & derivatives (cysteinyl leukotrienes)	Eosinophils, mast cells, macrophages, basophils	Provoke bronchoconstriction. Promote mucous production. Promote sensitivity to histamine. Increase permeability of vascular tissue to immune cells. Promote secretion of IL-4 and IL-5 by lymphocytes.

Leukotrienes (LTs) and the pathophysiology of allergic diseases.

Leukotrienes are inflammatory mediators (Brain and Williams, 1990; Samuelsson, 1983) that play a significant role in the pathophysiology of allergic diseases, including asthma, atopic dermatitis, allergic rhinitis and food allergy. Leukotrienes are derived from the fatty acid arachidonic acid (AA), which is concentrated in the membrane phospholipids of blood cells. There are two types of leukotrienes. Cysteinyl leukotrienes (LTC₄, LTD₄ and LTE₄) directly promote allergic inflammation in target tissues and increase the synthesis of other proinflammatory mediators, such as cytokines. The chemotactic leukotriene, LTB₄, is a chemoattractant for immune cells and activates neutrophils leading to endothelial leakage.

Key criteria (Busse 1996) that implicate LTs in the pathophysiology of AD, AS and AR include the following:

1. LTs are found in fluids removed from the involved tissues;
2. Tissues with receptors for LTs are found in the affected tissues;
3. The administration of exogenous LTs mimics the effects of AD, AS and AR in the relevant tissues;
4. Reducing LT activity, whether by inhibiting their synthesis or by blocking their receptors, ameliorates disease symptoms.

LTB₄ is produced primarily by neutrophils and monocyte/macrophages (O'Byrne 1997; Wenzel, 2003). The role of LTB₄ in allergic disease is multifactorial. First, LTB₄ is a chemoattractant for immune cells such as neutrophils, eosinophils and monocytes in the respiratory tract (Busse, 1996; Busse, 1998) and in skin (Bisgaard et al, 1986). These leukocytes are recruited from the general circulation to the site of inflammation in lungs, nasal tissues and skin. Second, LTB₄ activates neutrophils leading to endothelial leakage which further promotes allergic inflammation (Busse, 1998; Wenzel 2003). Finally, LTB₄ indirectly mediates pain (Bisgaard and Kristensen, 1985; Brain and Williams, 1990)

The roles of the cysteinyl leukotrienes, LTC₄, LTD₄ and LTE₄, are multifaceted (Bisgaard, 2001). When produced in lung tissue, they are powerful bronchoconstrictors; LTC₄, LTD₄ for example, are ca. 1000x more powerful than histamine (Bisgaard, 2001). Cysteinyl leukotrienes are produced by resident mast cells and alveolar macrophages, as well as by peripheral blood eosinophils (Bisgaard, 2001; Eapen and Busse, 2002). When produced in sinuses, cysteinyl leukotrienes promote the production of mucous, promote cell-mediated inflammation and promote synthesis of inflammatory cytokines (Bisgaard, 2001; Busse, 1998; Peters-Golden and Henderson, 2005). Airborne allergen exposure provokes leukotriene synthesis in nasal tissues (Busse, 1996). Cysteinyl leukotrienes also promote sensitivity to histamine by immune cells (Pynaert et al., 1999.) In skin, these leukotrienes increase the permeability of vascular tissue to inflammatory cells, and epidermal cells are sites of leukotriene synthesis (Rackal and Vender, 2004; Talbot et al., 1985).

In all of these allergic diseases, there is an overproduction of leukotrienes: the specific disease(s) present in any one individual may be determined in part by the location(s) of leukotriene synthesis.

Production of and response to leukotrienes by allergic individuals

Leukotrienes are important for innate immunity and the physiological response to pathogens (Peters-Golden and Coffey, 1999). However, excess production of leukotrienes promotes the inappropriate response to harmless antigenic substances that characterizes allergy.

Blood leukocytes of allergic individuals produce higher levels of leukotrienes upon stimulation than do the same types of cells from nonallergic individuals (Bisgaard, 2001; Sampson et al., 1992). Further, the response of allergic individuals to leukotrienes is exaggerated compared to the response by nonallergic individuals.

Leukotrienes in Atopic Dermatitis (AD).

Both LTB₄ and cysteinyl leukotrienes are implicated in the pathophysiology of AD (reviewed in Ruzicka, 1988, 1989). Cysteinyl leukotriene levels (urinary LTE₄) were significantly higher in patients with atopic dermatitis than in healthy volunteers (Fauler et al., 1993; Hishinuma et al. 2001). Others have documented increased levels of LTE₄ in urine from children with severe AD (Øymar et al., 2005) and from adults (Adamek-Guzik et al. 2002) during exacerbations but not during remission. Hon et al. (2004) found that disease severity as measured by SCORAD was significantly correlated with measures of urinary LTE₄ in children. In contrast, Sansom et al. (1997) reported no elevation in urinary LTE₄ levels in seven patients during and after a flare episode compared to a normal range from control subjects. LTB₄ can be isolated from skin lesions of atopic dermatitis (Fogh et al., 1989; Ikai and Imamura, 1993; Ruzicka et al., 1986). Increased production of LTB₄ *in vivo* has been reported in the skin of atopic patients after allergen specific challenge (Bisgaard et al., 1985; Koro et al., 1999).

Neuber et al. (1991) and Hilger et al. (1991) have reported that the spontaneous release of LTB₄ and LTC₄ from neutrophils is three times higher in patients with AD than in controls. Cysteinyl LT release from basophils (Shichijo et al., 1995) and eosinophils (Schauer et al., 1995) isolated from AD patients is increased compared to healthy controls. Enhanced spontaneous and stimulated releasability of LTC₄ from leukocytes of patients with AD compared with normal controls has been reported (Ruzicka 1988; Sansom et al. 1997). Elevated spontaneous release of LTC₄ from circulating basophils has been documented in severe compared to mild AD (James et al. 1993). Peripheral blood leukocytes from subjects with AD have a lower threshold of stimulus required for leukotriene release, and release higher levels of leukotrienes when stimulated than do leukocytes from normal subjects (Ruzicka and Ring, 1987). Shimizu et al., (1994) hypothesized that the increased production of LTC₄ *ex vivo* was due to eosinophilia rather than due to enhanced releasability of the leukotrienes in AD.

Since AD commonly occurs in patients with other atopic diseases such as asthma, it has been noted that patients treated with a leukotriene antagonist for their asthma symptoms also had some improvement in their skin condition. Case studies (Angelova-Fischer and Tsankov, 2005; Carucci et al., 1998) and randomized controlled studies (Capella et al., 2001; Eustachio et al., 2002; Rackal and Vender, 2004) in subjects with AD confirmed this observation.

The important role of the enzyme LTA₄ hydrolase, which synthesizes LTB₄, in the pathogenesis and development of atopic dermatitis was evaluated by Okano-Mitani et al. (1996). Their findings include the following; The LTA₄ hydrolase activities in the supernatant fraction of peripheral blood polymorphonuclear leukocytes (PMN) were significantly higher in preparations of cells from severe AD patients than in those from moderate and mild AD patients and normal controls. LTA₄ hydrolase activities were also significantly higher in peripheral blood mononuclear cells (PBMC) from severe AD patients than in those from moderate and mild AD patients and normal controls. LTA₄ hydrolase activities in PMN were reduced after improvement of the disease in eight patients with severe or moderate AD. The enhanced ability of neutrophils from AD

patients to produce LTB₄ from exogenous LTA₄ *in vitro* was recognized previously (Hilger et al., 1991). See also the review by Ikai and Imamura, 1993.

Leukotrienes in Asthma.

The cysteinyl leukotrienes are potent bronchoconstrictors produced by immune cells resident in lung tissue. These mediators are direct causal agents of many of the symptoms of asthma, including airway smooth muscle constriction, airway hyperresponsiveness, eosinophil migration, vascular permeability, and edema (Brain and Williams, 1990; Busse, 1998; Busse and Kraft, 2005; Donnelly et al. 1995; O'Byrne 1997; Wenzel, 2003). Leukocytes from asthmatics produce 3 – 5x higher levels of leukotrienes than do leukocytes from healthy controls (Sampson et al., 1992). The 5-LO enzyme may be upregulated in asthmatic subjects (Mita et al., 1985). Further, the response to inhaled leukotrienes is exaggerated in asthmatics compared to healthy subjects (O'Byrne 1997). In asthmatic children, concentrations of cysteinyl leukotrienes in exhaled breath condensates were significantly higher than in control children (Baraldi et al., 2003; Zanconato et al, 2004; Zsuzsanna et al., 2002). Asthmatic children with exercise-induced bronchoconstriction (EIB) had higher concentrations of cysteinyl leukotrienes in exhaled breath condensates than control children or asthmatic children without EIB (Carraro et al., 2005). Exaggerated production of cysteinyl leukotrienes is also evident in aspirin-induced asthma (Antczak et al., 2002).

Leukotrienes in Allergic Rhinitis (AR)

Leukotrienes mediate many of the typical symptoms of allergic rhinitis, including nasal congestion, mucous secretion and edema (Busse and Kraft, 2005; Peters-Golden and Henderson, 2005). CysLTs are overproduced by patients with allergic rhinitis within minutes of nasal allergen exposure (Peters-Golden and Henderson, 2005). Increased production of LTC₄ *in vivo* has been reported in the tear fluid (Bisgaard et al., 1985) and intact skin (Talbot et al., 1985) of atopic patients after allergen specific challenge. These lipid mediators interact with receptors, particularly the cysLT1 receptor, on eosinophils, mast cells, macrophages, and neutrophils (Steinke and Borish, 2004). They also stimulate production of other proinflammatory mediators, such as cytokines IL-4 and IL-5 (Steinke and Borish, 2004).

Peters-Golden and Henderson (2005) recently reviewed the significant role that leukotrienes play in allergic rhinitis. “Many of the cells involved in the pathophysiology of allergic rhinitis produce and release CysLTs. This production and release begin early in the allergic response, since unlike cytokines, which require transcription and translation before synthesis, all the enzymes necessary to produce CysLTs are already present in inflammatory cells. During the early phase response, mast cells and basophils are the primary source of CysLTs, which act both locally and systemically to stimulate the production, recruitment, and activation of additional inflammatory cells. The newly recruited inflammatory cells, predominantly eosinophils but also monocytes and macrophages, are the primary source of CysLTs during the late-phase reaction”.

Strategies to reduce the impact of leukotrienes in allergic diseases.

Two different types of LT-modulating agents are 5-lipoxygenase inhibitors and LT receptor antagonists (LTRAs). Since the 5-lipoxygenase inhibitor acts at the initial step in the LT synthetic pathway, it has the ability to alter the production of all the leukotrienes (Chari et al., 2001; Julemont et al., 2003), including LTB₄, while the receptor antagonists target only the cysteinyl LTs, LTC₄, LTD₄, and LTE₄. There are several ways to assess leukotriene synthesis *in vivo* that have been used in clinical trials: LTB₄ production by stimulated whole blood *ex vivo* (e.g. Rubin et al., 1991; Surette et al, 2003b), urinary LTE₄ (e.g. Israel et al, 1993), and LTs in exhaled breath condensate (e.g. Baraldi et al., 2003; Zsuzsanna et al., 2002).

LT modulating agents have been assessed as monotherapy as well as in combination with other therapies. In most cases, LT modifying agents have a clinically significant benefit in combination with other therapies.

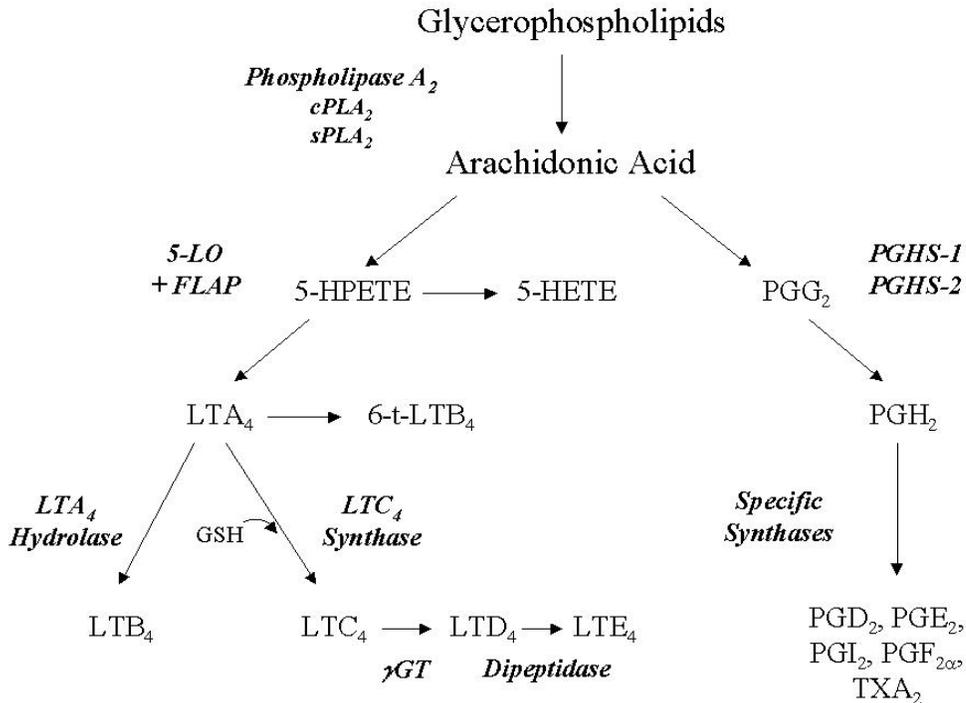


Fig. 1. Leukotriene (LT) biosynthetic pathway (left side of Figure). The enzyme 5-lipoxygenase (5-LO) acts upon free arachidonic acid as the first step in the pathway. The 5-lipoxygenase inhibitor, zileuton, as well as metabolites from the medical food EFFICAS CARE, act to inhibit 5-LO and thus blunt the entire leukotriene pathway. The leukotriene receptor antagonists, e.g. montelukast, do not impact synthesis of leukotrienes, but block the activity of the cysteinyl leukotrienes at the receptor.

Leukotriene Modulation in Atopic Dermatitis

Evidence in the literature provides a pathophysiological rationale for the use of cysLT receptor blockers in the treatment of AD (reviewed in Kagi, 2001). Case reports confirm that LTRs may reduce symptoms of AD even in severe cases (Zabawski et al., 1999) although an LTR had no significant impact on moderate to severe AD in one controlled trial (Veien et al., 2005). In addition, leukotriene synthesis inhibition would likely be beneficial, given the role of LTB₄ in AD. This topic was recently reviewed by Rackal and Vender (2004).

Impacts of 5-LO inhibition in atopic dermatitis include:

1. Significant reduction in disease dissatisfaction (p=0.03) (Woodmansee and Simon, 1999).
2. Significant reduction in objective skin scores (p=0.03) (Woodmansee and Simon, 1999).

Impacts of LTRAs in atopic dermatitis include:

1. Reduced body surface affected (Carucci et al., 1998)
2. Reduced severity of AD (Biswas et al., 2001; Capella et al., 2001; Friedman et al., 2000; Galindo et al., 2000; Pei et al., 2001; Yanase and David-Bajar, 2001; Zabawski et al., 1999).
3. Reduced itch (Capella et al., 2001; Friedman et al., 2000)
4. Steroid sparing (Carucci et al., 1998; Yanase and David-Bajar, 2001; Zabawski et al., 1999)

LTRAs have also been reported to reduce severity of urticaria (Erbagci 2002)

Leukotriene Modulation in Asthma

Impacts of 5-LO inhibition in asthma include:

1. Significant improvement in quality of life and asthma control in adult subjects (Israel et al., 1996).
2. Improved FEV₁ (Liu et al., 1996)
3. Reduced β -agonist use (Liu et al., 1996)
4. Reduced cold-air induced bronchoconstriction (reviewed in Bisgaard, 2001)
5. Reduced eosinophilia (reviewed in Bisgaard, 2001; Liu et al., 1996)
6. Reduced exercise-induced bronchoconstriction (reviewed in Horwitz et al., 1998)
7. 50% reduction in emergency room visits and hospitalizations (reviewed in Wenzel, 1988).

Impacts of LTRAs in asthma include:

1. Improved pulmonary function in adult patients with asthma (Barnes et al 1997; Barnes et al. 2001, Fish et al, 1997; Kemp et al. 1999; Reiss et al. 1998).
2. Significantly improved pulmonary function and reduced symptoms in children, irrespective of concomitant inhaled corticosteroid (ICS) use (Knorr et al. 1998; Knorr et al. 2001; Pearlman et al. 2000).

3. Significant tapering of ICS dose without loss of asthma control (Lofdahl et al. 1999).
4. Significantly reduced percentage of days with daytime asthma symptoms and reduced nighttime awakenings in children (Knorr et al. 2001; Pearlman et al. 2000).
5. Reduced airway hyperresponsiveness (Carratu et al. 2003; Rorke et al. 2002; Westbroek and Pasma, 2000; Yoshida et al. 2000).
6. Decreased daytime asthma symptoms and nocturnal awakenings in adult patients (Reiss et al. 1998).
7. Significantly improved the asthma quality of life in all four domains (activity, symptoms, emotional function, and exposure to environmental stimuli) (Reiss et al. 1998; Nathan et al. 1998).
8. Decreased eosinophil levels in blood, BAL fluid, and sputum of adults (Calhoun et al. 1998; Pizzichini et al. 1999; Reiss et al. 1998) and decreased eosinophil levels in blood of children (Knorr et al. 1998, 2001; Stelmach et al. 2002).
9. Significant reduction in the decreased pulmonary function after exercise in asthmatics with exercise-induced bronchoconstriction (Dahlen et al. 2002; Kemp et al. 1998; Pearlman et al. 1999).

Leukotriene Modulation in Allergic Rhinitis

Nasal congestion was reduced by inhibition of 5-LO (Knapp and Murray, 1994).

Impacts of LTRAs in allergic rhinitis include:

- Reduced rhinorrhea, nasal itching and nasal obstruction (Jiang, 2006).
- Reduced daytime nasal symptoms (congestion, rhinorrhea, pruritus, and sneezing) and nighttime symptoms (difficulty sleeping, nighttime awakenings, and congestion on awakening) (Patel et al., 2005)
- Reduced daytime nasal symptoms (congestion, rhinorrhea, pruritus, and sneezing), nighttime symptoms (difficulty sleeping, nighttime awakenings, and congestion on awakening) and daytime eye symptoms (Nayak et al., 2002; Philip et al. 2002; van Adelsberg et al. 2003)
- Significantly reduced daytime nasal symptoms, daytime eye symptoms, nighttime symptoms and composite symptom score when combined with loratidine compared to placebo (Meltzer et al., 2000; Nayak et al., 2002)
- Significantly improved rhinoconjunctivitis-specific quality-of-life score, which included significant improvement in domains such as activity, ocular symptoms, practical problems, and emotional symptoms (Meltzer et al., 2000; Nayak et al. 2002; Patel et al., 2005; Philip et al. 2002; van Adelsberg et al. 2003).
- Decreased peripheral blood eosinophils (Philip et al., 2002)
- Decreased number of eosinophils in nasal lavage fluid of patients with both asthma and allergic rhinitis (Piatti et al. 2003).
- Reduced asthma and allergic rhinitis symptoms in patients with both asthma and allergic rhinitis (Piatti et al. 2003).

Leukotrienes and Quality of Life

The reduction of leukotriene levels in subjects with asthma (Biernacki et al., 2005; Busse and Kraft, 2005; Israel et al., 1996; Nathan et al., 1998), atopic dermatitis (Woodmansee and Simon, 1999), and allergic rhinitis (Patel et al., 2005) has been reported to improve quality of life.

EFFICAS CARE: Mechanisms of action relevant to allergic diseases.

EFFICAS CARE is a medical food that reduces production of leukotrienes by inhibiting the action of the 5-lipoxygenase enzyme. The role of this enzyme in leukotriene synthesis is illustrated in Figure 1, above.

Leukotriene Production and the Arachidonic Acid Pathway.

When immune cells are stimulated, fatty acids are cleaved from the membrane phospholipids by phospholipase¹. Leukotrienes are formed from arachidonic acid (AA), in response to a stimulus such as allergen exposure. The free AA is a substrate for the enzyme 5-Lipoxygenase (5-LO)², the first enzyme in the pathway for synthesis of the bronchoconstricting cysteinyl leukotrienes, LTC₄, LTD₄, LTE₄, and the chemotactic leukotriene LTB₄ (Bisgaard, 2001). 5-LO produces a short-lived intermediate from AA, 5-HPETE. A synthase then produces LTA₄, another short lived intermediate which is a branch point in the leukotriene pathway. This intermediate is a substrate for LTA₄ hydrolase, producing LTB₄. The intermediate may also combine with a glutathione residue to produce LTC₄. Further processing of LTC₄ produces LTD₄ and LTE₄. Inhibition of the first enzyme in the pathway, 5-LO, reduces levels of all the leukotrienes (Julemont et al., 2003).

The same stimulus that releases AA from membranes will release other highly unsaturated fatty acids, such as DGLA and EPA. DGLA decreased LTB₄ production by human mononuclear leukocytes *in vitro* in a dose-dependent manner (Iverson et al., 1992). Unlike arachidonic acid and EPA, DGLA is not a substrate for 5-LO, but instead is converted by 15-LO to 15-OH-DGLA (aka 15-HETrE). 15-HETrE blocks conversion of AA to LTA₄ by direct inhibition of 5-LO (Guichardant et al., 1993) and is an even stronger inhibitor of LTB₄ production than is DGLA (50x). Inhibition of 5-LO activity in human neutrophils *ex vivo* has also been demonstrated in cells supplemented with GLA or DGLA (Chilton-Lopez et al., 1996). Neutrophils metabolize DGLA to 15-HETrE (Chilton-Lopez et al., 1996). Dietary GLA increases DGLA in phospholipids and decreases *ex vivo* elaboration of LTB₄ in healthy human volunteers (Surette et al., 2003a; Ziboh and Fletcher, 1992).

When EPA is liberated from membrane phospholipid, it competes with arachidonic acid for the 5-LO enzyme (Lands, 1992). Dietary EPA (provided in fish oil) has been demonstrated in multiple studies to actively reduce levels of AA in membrane

¹ Inhibition of phospholipase is the primary anti-inflammatory activity of corticosteroids.

² Arachidonic acid is a substrate for numerous enzymes that influence inflammation. For brevity, only the leukotriene pathway is introduced here.

phospholipids (Chapkin et al., 1991; Miller et al., 1991; Stulnig, 2003). Dietary EPA reduced AA levels in murine macrophages (Chapkin et al., 1991) but not in human neutrophils (Chilton et al., 1993). EPA inhibits the enzyme delta-5 desaturase that would otherwise produce AA from DGLA (Nassar et al., 1986), so that higher doses of GLA can be provided safely in a medical food as long as EPA is also provided in sufficient quantity. Finally, in contrast to DGLA, EPA is a substrate for 5-LO, and thus competes with AA for the enzyme's active sites. 5-series leukotrienes are produced from the action of 5-LO on EPA, but these have reduced bronchoconstricting activity compared to LTB₄, LTC₄, LTD₄ and LTE₄ (Schwartz, 2000). Peritoneal macrophages from mice fed fish oils produced fewer leukotrienes overall than those from control mice, and the ratio of series 4 to series 5 leukotrienes was dramatically altered in favor of series 5.

Clinical Studies Demonstrating Impact of the Medical Food on Leukotrienes

EFFICAS CARE, a medical food formulation containing specified amounts and ratios of GLA and EPA, has been demonstrated to significantly reduce leukotriene synthesis (LTB₄) by stimulated whole blood in healthy subjects and asthmatic subjects (Surette et al., 2003a,b).

Two preliminary clinical trials with the active ingredients were conducted to determine the minimum effective levels needed to reduce leukotriene biosynthesis and prevent increases in plasma arachidonic acid (Barham et al., 2000; Surette et al., 2003a). These were followed by three clinical studies conducted to evaluate the safety, tolerability and effectiveness of the formulated medical food product. These studies were: a single-center, randomized, double-blind, placebo-controlled, parallel-group, escalating-intake inpatient trial in healthy adult subjects (Surette et al., 2003a); a pharmacokinetic study in asthmatic children and adolescents (Efficas, unpublished); and a safety and efficacy study in adult asthmatics (Surette et al., 2003b). Results in these studies were consistent. The bioavailability of the bioactive fatty acids was significantly improved by formulating them into the emulsion product. It was also convincingly demonstrated that the product was well-tolerated and was effective in lowering the production of leukotrienes in healthy subjects and in those with asthma. The emulsified medical food providing 750 mg GLA + 500 mg EPA significantly reduced ($p < 0.05$) *ex vivo* whole blood leukotriene synthesis by 23% in asthmatic subjects (Surette et al., 2003b).

Proven Leukotriene Reduction in Healthy Subjects

In a placebo-controlled clinical study involving 47 healthy adults, Efficas Care significantly decreased leukotriene production within 14 days (Surette et al., 2003a). The clinical study also demonstrated that the medical food was well tolerated in this population.

Leukotriene (LTB₄) Reduction ($p < 0.03$)

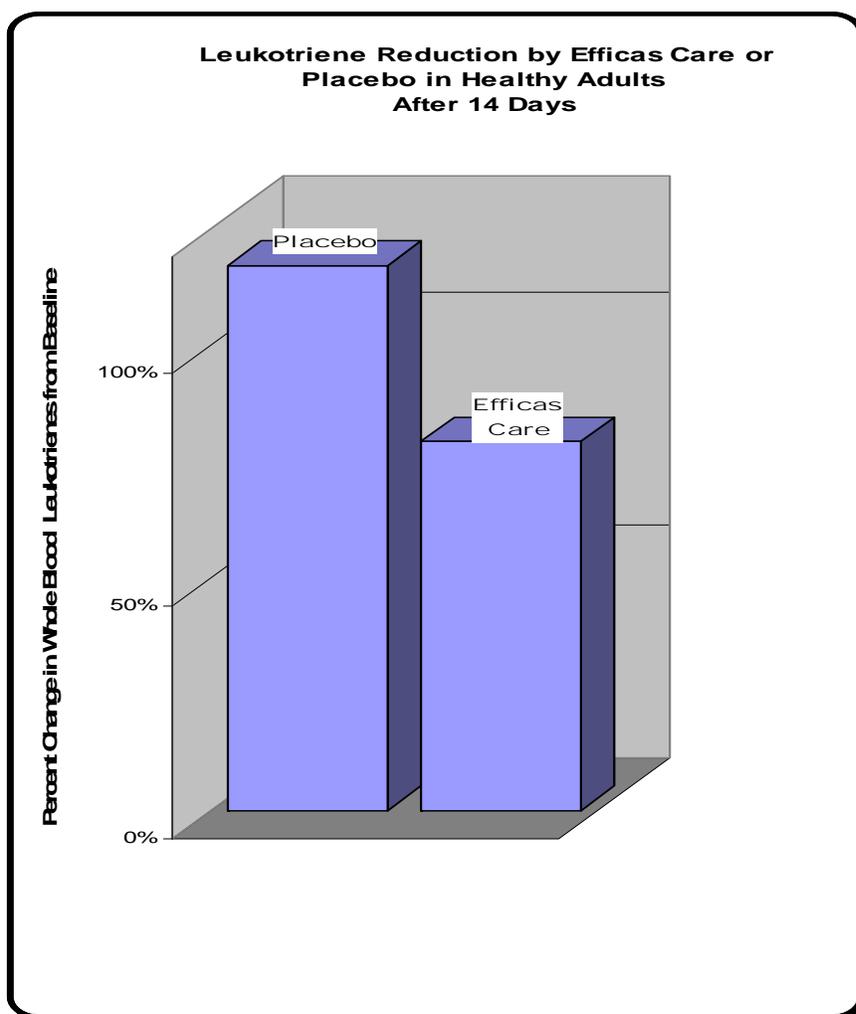
Efficas Care vs placebo – mean change from baseline
-4.1 vs. 3.8 ng/ml per 10⁶ neutrophils ($p < 0.03$)*

Whole blood leukotriene (LTB₄) concentrations (ng • ml⁻¹ • 10⁶ polymorphonuclear neutrophils) in healthy adults consuming Efficas Care or placebo for 2 weeks. Values are mean (SD).

	Baseline (ng x ml ⁻¹ x 10 ⁶ PMN)	Day 14 (ng x ml ⁻¹ x 10 ⁶ PMN)
Group	Mean (SD)	Mean (SD)
Placebo	22.2 (7.3)	26.0 (16.2)
10g Efficas Care TM	19.8 (4.8)	15.7* (6.0)

*Significantly different compared to placebo determined by ANCOVA, p<0.03.

SD = standard deviation; PMN = polymorphonuclear neutrophils.



Proven Leukotriene Reduction in Asthmatic Patients

In a placebo-controlled clinical study involving 43 adult patients with mild to moderate atopic asthma, Efficas Care significantly decreased leukotriene production within 28 days (Surette et al., 2003b). The clinical study also demonstrated that the medical food was well tolerated in this population.

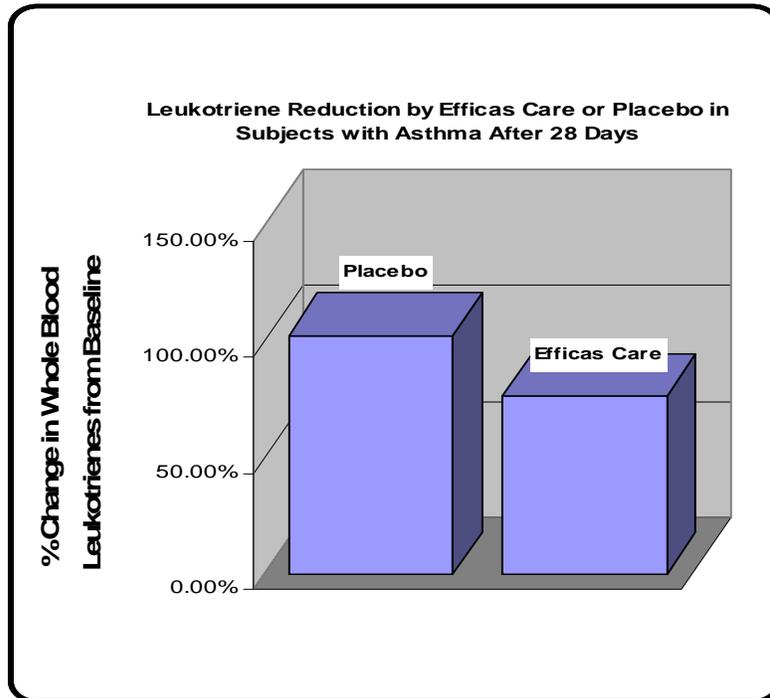
Leukotriene (LTB₄) Reduction (p< 0.05)

Efficas Care vs placebo – mean change from baseline
-4.1 vs. 0.6 ng/ml per 10⁶ neutrophils (p< 0.05)*

Whole blood leukotriene (LTB₄) concentrations (ng • ml⁻¹ • 10⁶ Polymorphonuclear neutrophils) in patients with atopic asthma consuming Efficas Care or placebo for 4 weeks. Values are mean (SE).

	Baseline (ng x ml ⁻¹ x10 ⁶ PMN)	Day 28 (ng x ml ⁻¹ x10 ⁶ PMN)
Group	Mean (SE)	Mean (SE)
Placebo	18.8±3.9	19.4±2.6
10g Efficas Care™	17.6±3.1	13.5±1.5 *

*Significantly different compared to placebo determined by ANCOVA (p<0.05).



Studies Demonstrating Impact of the Medical Food on Quality of Life and Disease Management

Patient assessments of their symptoms are an important measure of benefit (quality of life). The medical food improved subjective assessment of asthma in 8 of 11 patients (72%) versus 37.5% of subjects on placebo after 4 weeks of use (Surette et al., manuscript in preparation). An additional open label consumer study evaluated the impact on QOL of the medical food in subjects with asthma, allergic rhinitis or eczema. Mean scores for the validated QOL instruments MiniAQLQ and ACQ improved significantly ($p < 0.001$) in subjects with asthma (Surette et al., manuscript in preparation). QOL scores also improved significantly ($p < 0.001$) in subjects with allergic rhinitis and eczema (Efficas, unpublished).

Conclusion:

EFFICAS CARE has been shown in clinical trials to alter the pools of fatty acid precursors of leukotrienes, to significantly reduce leukotrienes in healthy individuals as well as in asthmatics and to improve the quality of life in subjects with asthma, allergic rhinitis and eczema. Leukotrienes contribute significantly to the pathophysiology of allergic diseases, including asthma, allergic rhinitis and eczema. The medical food, EFFICAS CARE, blunts leukotriene synthesis via a nutritional mechanism, and thus is suitable for use in multiple allergic diseases.

Literature Cited

- Adamek-Guzik T et al., 2002. Urinary leukotriene levels are increased during exacerbation of atopic eczema/dermatitis syndrome. Relation to clinical status. *Allergy* 57: 732-736.
- Angelova-Fischer I and Tsankov N. 2005. Successful treatment of severe atopic dermatitis with cysteinyl leukotriene receptor antagonist montelukast. *Acta Dermatoven APA* 14:115-119.
- Antczak, A. et al. 2002. Increased Exhaled Cysteinyl-Leukotrienes and 8-Isoprostane in Aspirin-Induced Asthma. *Am J Respir Crit Care Med* 166: 301-306
- Baraldi, E., et al. 2003. Cysteinyl Leukotrienes and 8-Isoprostane in Exhaled Breath Condensate of Children with Asthma Exacerbations. *Thorax* 58:505-509.
- Barham JB et al., 2000. Addition of eicosapentaenoic acid to gamma-linolenic acid-supplemented diets prevents serum arachidonic acid accumulation in humans. *J Nutr* 130: 1925-1931.
- Barnes NC et al. 1997. Pranlukast, a novel leukotriene receptor antagonist: results of the first European, placebo controlled, multicentre clinical study in asthma. *Thorax* 52:523.
- Barnes N et al. 2001. Analysis of montelukast in mild persistent asthmatic patients with near-normal lung function. *Respir Med* 95:379.
- Biernacki, W.A. et al. 2005. Effect of Montelukast on Exhaled Leukotrienes and Quality of Life in Asthmatic Patients. *CHEST* 128:1958-1963.
- Bisgaard H. and Kristensen JK 1985. Leukotriene B₄ produces hyperalgesia in humans. *Prostaglandins* 30: 791-797.
- Bisgaard H. et al. 1985. Production of leukotrienes in human skin and conjunctival mucosa after specific allergen challenge. *Allergy* 40: 417-423.
- Bisgaard H. et al, 1986. Chemotactic activity of LTB₄ in man. *Allergy* 41: 365-372.
- Bisgaard, H. 2001. Leukotriene Modifiers in Pediatric Asthma Management. *Pediatrics* 107:381-390.
- Biswas P et al. 2001. Montelukast and improvement of eczema: observations from a prescription event monitoring study in England. *Int J Clin Pharmacol Ther* 39: 529-533.
- Borish LC and Steinke JW. 2003. Cytokines and chemokines. *J Allergy Clin Immunol* 111:S460-S475.

- Brain SD and Williams TJ. 1990. Leukotrienes and inflammation. *Pharmacol Ther* 46:57-66.
- Busse, WW. 1996. The role of leukotrienes in asthma and allergic rhinitis. *Clin Exp Allergy* 26: 868-879.
- Busse WW. 1998. Leukotrienes and inflammation. *Am J Respir Crit Care Med* 157 (6 Pt 2): S210-3.
- Busse WW and Kraft M. 2005. Cysteinyl leukotrienes in allergic inflammation. *Chest*. 127:1312.
- Calhoun WJ et al. 1998. Effect of zafirlukast (Accolate) on cellular mediators of inflammation: bronchoalveolar lavage fluid findings after segmental antigen challenge. *Am J Respir Crit Care Med* 157:1381.
- Capella GL et al. 2001. A randomized trial of leukotriene receptor antagonist montelukast in moderate-to-severe atopic dermatitis of adults. *Eur J Dermatol* 11(3): 209-213.
- Carraro SC et al. 2005. Exhaled breath condensate cysteinyl leukotrienes are increased in children with exercised-induced bronchoconstriction. *J Allergy Clin Immunol* 115: 764-770.
- Carratu P et al. 2003. Effect of zafirlukast on methacholine and ultrasonically nebulized distilled water challenge in patients with mild asthma. *Respiration* 70:249.
- Carucci JA et al. 1998. The leukotriene antagonist zafirlukast as a therapeutic agent for atopic dermatitis. *Arch Dermatol* 134:785-6.
- Chapkin, R.S., Akoh, C.C. and Miller C.G. 1991. Influence of dietary n-3 fatty acids on macrophage glycerophospholipid molecular species and peptidoleukotriene synthesis. *J Lipid Res* 32: 1205-1213.
- Chari S et al. 2001. A role for leukotriene antagonists in atopic dermatitis. *Am J Clin Dermatol* 2:1-6
- Chilton, F.H., et al. 1993. Dietary n-3 Fatty Acid Effects on Neutrophil Lipid Composition and Mediator Production. *J Clin Invest* 91:115-122.
- Chilton-Lopez T, Surette ME, Swan DD, Fonteh AN, Johnson MM and Chilton FH. 1996. Metabolism of gammalinolenic acid in human neutrophils. *J Immunol* 156:2941-2947.

- Dahlen B et al. 2002. Influence of zafirlukast and loratadine on exercise-induced bronchoconstriction. *J Allergy Clin Immunol* 109:789.
- Donnelly AL et al. 1995. The leukotriene D₄- receptor antagonist, ICI 204,219, relieves symptoms of acute seasonal allergic rhinitis. *Am J Respir Crit Care Med* 151(6):1734.
- Eapen S.S. and Busse W.W. 2002. Asthma. In: Zweiman, B. and Schwartz, L.B. eds. *Inflammatory Mechanisms in Allergic Disease*. Marcel Dekker, New York.
- Erbagci Z. 2002. The leukotriene receptor antagonist montelukast in the treatment of chronic idiopathic urticaria: A single-blind, placebo controlled, crossover clinical study. *J Allergy Clin Immunol* 110(3):484.
- Eustachio N et al. 2002. Efficacy and tolerability of montelukast as a therapeutic agent for severe atopic dermatitis in adults. *Acta Derm Venereol* 82(4): 297-298.
- Fauler J et al., 1993. Enhanced synthesis of cysteinyl leukotrienes in atopic dermatitis. *Br J Dermatol* 128: 627-630.
- Fish JE et al. 1997. Zafirlukast for symptomatic mild-to-moderate asthma: a 13-week multicenter study. The Zafirlukast Trialists Group. *Clin Ther* 19:675.
- Fogh K, Herlin T, Kragballe K. 1989. Eicosanoids in skin of patients with atopic dermatitis: prostaglandin E₂ and leukotriene B₄ are present in biologically active concentrations. *J Allergy Clin Immunol* 83 : 450.
- Friedmann PS et al., 2000. Treatment of atopic dermatitis with montelukast (Abstr). *Allergy Clin Immunol Int* 2000; suppl 2: 56.
- Galindo G et al., 2000. Use of montelukast in the treatment of atopic dermatitis: A case report (Abstr). *Ann Allergy Asthma Immunol* 84: 154.
- Guichardant M et al., 1993. Stearidonic acid, an inhibitor of the 5-lipoxygenase pathway. A comparison with timnodonic and dihomogammalinolenic acid. *Lipids* 28: 321-324.
- Hilger RA, Neuber K, König W. 1991. Conversion of leukotriene A₄ by neutrophils and platelets from patients with atopic dermatitis. *Immunology* 74 : 689.
- Hishinuma T et al. 2001. Increased urinary leukotriene E₄ excretion in patients with atopic dermatitis. *Br J Dermatol* 144(1):19-23.
- Holgate ST. 2000. The role of mast cells and basophils in inflammation. *Clin Exp Allergy* 30: 28-32.

- Hon KLE et al., 2004. Urinary leukotriene E₄ correlates with severity of atopic dermatitis in children. *Clin Exp Dermatol* 29: 277-281.
- Horrobin, D.F. 2000. Essential fatty acid metabolism and its modification in atopic eczema. *Am J Clin Nutr* 71 (suppl): 367S-372S.
- Horwitz RJ, McGill KA, Busse WW. 1998. The role of leukotriene modifiers in the treatment of asthma. *Am J Respir Crit Care Med* 157: 1363-71.
- Iikura M, et al., 2005. 5-lipoxygenase products regulate basophil functions: 5-Oxo-ETE elicits migration, and leukotriene B₄ induces degranulation. *J Allergy Clin Immunol* 116:578-585.
- Ikai K and Imamura S. 1993. Role of eicosanoids in the pathogenesis of atopic dermatitis. *Prostaglandins Leukot Essent Fatty Acids* 48 : 409-416.
- Israel E. et al., 1993. The effect of inhibition of 5-lipoxygenase by zileuton in mild-to-moderate asthma. *Ann Intern Med* 119: 1059-1066.
- Israel E. et al., 1996. Effect of treatment with zileuton, a 5-lipoxygenase inhibitor, in patients with asthma. A randomized controlled trial. Zileuton Clinical Trial Group. *JAMA* 275: 931-936.
- Iverson L et al. 1992. Effects of dihomogammalinolenic acid and 15-lipoxygenase metabolite on eicosanoid metabolism by human mononuclear leukocytes in vitro: selective inhibition of the 15-LO pathway. *Arch Dermatol Res* 284:222
- James JM et al. 1993. Patients with severe atopic dermatitis have activated circulating basophils. *J Allergy Clin Immunol* 91(6):1155.
- Jiang RS. 2006. Efficacy of a leukotriene receptor antagonist in the treatment of perennial allergic rhinitis. *J Otolaryngol* 35:117-121.
- Johnson MM et al. 1997. Dietary supplementation with gamma-linolenic acid alters fatty acid content and eicosanoid production in healthy humans. *J Nutr.* 127: 1435-1444.
- Julemont F. et al. 2003. Recent Developments in 5-Lipoxygenase Inhibitors. *Expert Opinion Ther. Patents* 13(1):1-13.
- Kagi MK. 2001. Leukotriene receptor antagonists – A novel therapeutic approach in atopic dermatitis? *Dermatology* 203:280.
- Kemp JP et al. 1998. Montelukast once daily inhibits exercise-induced bronchoconstriction in 6- to 14-year-old children with asthma. *J Pediatr* 133:424.

- Kemp JP et al. 1999. Therapeutic effect of zafirlukast as monotherapy in steroid-naive patients with severe persistent asthma. *Chest* 115:336.
- Knapp HR and Murray JJ. 1994. Leukotrienes as mediators of nasal inflammation. *Adv Prost Thromb Leuk Res* 22: 279-288.
- Knorr B et al. 1998. Montelukast for chronic asthma in 6- to 14-year-old children: a randomized, double-blind trial. Pediatric Montelukast Study Group. *JAMA* 279:1181.
- Knorr B et al. 2001. Montelukast, a leukotriene receptor antagonist, for the treatment of persistent asthma in children aged 2 to 5 years. *Pediatrics* 108:E48.
- Koro O et al. 1999. Chemical mediators in atopic dermatitis: involvement of leukotriene B₄ released by a type I allergic reaction in the pathogenesis of atopic dermatitis. *J Allergy Clin Immunol* 103: 663.
- Lands, W.E.M. 1992. Biochemistry and physiology of n-3 fatty acids. *FASEB J* 6: 2530-2536.
- Leung D et al. 2004. New insights into atopic dermatitis. *J Clin Invest* 113:651.
- Liu MC et al., 1996. Acute and chronic effects of a 5-lipoxygenase inhibitor in asthma: A 6-month randomized multicenter trial. *J Allergy Clin Immunol* 98: 859-71.
- Lofdahl C-G et al. 1999. Randomised, placebo controlled trial of effect of a leukotriene receptor antagonist, montelukast, on tapering inhaled corticosteroids in asthmatic patients. *BMJ* 319:8.
- Meltzer EO. 2000. Role for cysteinyl leukotriene receptor antagonist therapy in asthma and their potential role in allergic rhinitis based on the concept of "one linked airway disease". *Ann Allergy Asthma Immunol* 84:176-187.
- Meltzer EO, et al. 2000. Concomitant montelukast and loratidine as treatment for seasonal allergic rhinitis: A randomized, placebo-controlled clinical trial. *J Allergy Clin Immunol* 105: 917-922.
- Miller, C.C. et al. 1991. Dietary supplementation with ethyl ester concentrates of fish oil (n-3) and borage oil (n-6) polyunsaturated fatty acids induces epidermal generation of local putative anti-inflammatory metabolites. *J Invest Dermatol* 96: 98-103.
- Miller CC and Ziboh VA. 1988. Gammalinolenic acid-enriched diet alters cutaneous eicosanoids. *Biochem Biophys Res Comm* 154: 967-974.
- Mita H, Yui Y, Taniguchi N, Yasueda H, Shida T. 1985. Increased activity of 5-lipoxygenase in polymorphonuclear leukocytes from asthmatic patients. *Life Sci* 37:907-914

Nassar BA et al., 1986. The influence of dietary manipulation with n-3 and n-6 fatty acids on liver and plasma phospholipid fatty acids in rats. *Lipids* 21: 652-656.

Nathan RA et al. 1998. Zafirlukast improves asthma symptoms and quality of life in patients with moderate reversible airflow obstruction. *J Allergy Clin Immunol* 102:935-942.

Nayak AS et al. 2002. Efficacy and tolerability of montelukast alone or in combination with loratadine in seasonal allergic rhinitis: a multicenter, randomized, double-blind, placebo-controlled trial performed in the fall. *Ann Allergy Asthma Immunol* 88:592.

Neuber K, Hilger RA, König W. 1991. Interleukin-3, interleukin-8, FMLP and C5a enhance the release of leukotrienes from neutrophils of patients with atopic dermatitis. *Immunology* 73 :83–87

O’Byrne PM. 1997. Leukotrienes in pathogenesis of asthma. *Chest* 111(2 Suppl.): 27s.

Oettgen HC and Geha RS. 1999. IgE in asthma and atopy: cellular and molecular connections. *J Clin Invest* 104:829-835.

Okano-Mitani K et al. 1996. Leukotriene A₄ hydrolase in peripheral leukocytes of patients with atopic dermatitis. *Arch Dermatol Res* 288: 168-172.

Øymar K et al. 2005. Increased levels of urinary leukotriene E₄ in children with severe atopic eczema/dermatitis syndrome. *Allergy* 60:86.

Patel P et al. 2005. Randomized, double-blind, placebo-controlled study of montelukast for treating perennial allergic rhinitis. *Ann Allergy Asthma Immunol* 95(6): 551-557.

Pearlman DS et al., 1999. The leukotriene D₄-receptor antagonist Zafirlukast attenuates exercise-induced bronchoconstriction in children. *J Pediatr* 134:273-279.

Pearlman DS et al. 2000. Effectiveness and tolerability of zafirlukast for the treatment of asthma in children. *Clin Ther* 22:732.

Pei AY et al., 2001. Montelukast in the treatment of children with moderate-to-severe atopic dermatitis: a pilot study. *Ped Allergy Immunol* 12: 154-158.

Peters-Golden, M. and Coffey, M. 1999. Role of Leukotrienes in Antimicrobial Host Defense of Lung, *Clinical Reviews in Allergy and Immunology* 17:261-269.

Peters-Golden M and Henderson WR Jr. 2005. The role of leukotrienes in allergic rhinitis. *Ann Allergy Asthma Immunol* 94(6):609-618.

Philip G et al. 2002. Montelukast for treating seasonal allergic rhinitis: a randomized, doubleblind, placebo-controlled trial performed in the spring. *Clin Exp Allergy* 32:1020

Piatti G et al. 2003. Effects of zafirlukast on bronchial asthma and allergic rhinitis. *Pharmacol Res* 47:541.

Pizzichini E et al. 1999. Montelukast reduces airway eosinophilic inflammation in asthma: a randomized, controlled trial. *Eur Respir J* 14:12.

Prussin C and Metcalfe DD. 2003. IgE, mast cells, basophils, and eosinophils. *J Allergy Clin Immunol.* 111:S486.

Pynaert G. et al. 1999. Cysteinyl leukotrienes mediate histamine hypersensitivity ex vivo by increasing histamine receptor numbers. *Mol Med* 5(10): 685-692.

Rachelefsky G. 1997. Childhood asthma and allergic rhinitis: the role of leukotrienes. *J Pediatr* 131: 348-355.

Rackal JM and Vender RB. 2004. The treatment of atopic dermatitis and other dermatoses with leukotriene antagonists. *Skin Therapy Lett.* 9:1-5

Reiss TF et al. 1998. Montelukast, a once-daily leukotriene receptor antagonist, in the treatment of chronic asthma: a multicenter, randomized, double-blind trial. Montelukast Clinical Research Study Group. *Arch Intern Med* 158:1213.

Rorke S et al. 2002. Role of cysteinyl leukotrienes in adenosine 5-monophosphate induced bronchoconstriction in asthma. *Thorax* 57:323.

Rubin P et al., 1991. Pharmacokinetics, safety, and ability to diminish leukotriene synthesis by zileuton, an inhibitor of 5-lipoxygenase. *Agents Actions Suppl.* 35: 103-116.

Ruzicka T 1988. The physiology and pathophysiology of eicosanoids in the skin. *Eicosanoids* 1:59-72.

Ruzicka T 1989. Leukotrienes in atopic eczema. *Acta Derm Venereol Suppl* 144: 48-49.

Ruzicka T et al. 1986. Skin levels of arachidonic acid-derived inflammatory mediators and histamine in atopic dermatitis and psoriasis. *J Invest Dermatol* 86:105-108.

Ruzicka T and Ring J. 1987. Enhanced releasability of prostaglandin E₂ and leukotrienes B₄ and C₄ from leukocytes of patients with atopic eczema. *Acta Derm Venereol* 67(6):469-475.

Ruzika et al., 1986. Skin levels of arachidonic acid-derived inflammatory mediators and histamine in atopic dermatitis. *J Invest Dermatol* 86: 105-108.

- Sampson AP et al. 1992. Enhanced leukotriene synthesis in leukocytes of atopic and asthmatic subjects. *Br J Clin Pharmacol* 33:423.
- Samuelsson B 1983. Leukotrienes: mediators of immediate hypersensitivity reactions and inflammation. *Science* 220: 568-575.
- Sansom JE et al. 1997. Urinary leukotriene E₄ levels in patients with atopic dermatitis. *Br J Dermatol* 136:790.
- Schauer U et al., 1995. Blood eosinophils, eosinophil-derived proteins, and leukotriene C₄ generation in relation to bronchial hyperreactivity in children with atopic dermatitis. *Allergy* 50: 126-132.
- Schwartz, J. 2000. Role of Polyunsaturated Fatty Acids in Lung Disease. *Am J Clin Nutr* 71:393S-396S.
- Semper A et al. 2003. Surface expression of FcεpsilonRI on Langerhans' cells of clinically uninvolved skin is associated with disease activity in atopic dermatitis, allergic asthma, and rhinitis. *J Allergy Clin Immunol* 112:411.
- Shichijo M et al., 1995. Relationship between histamine release and leukotrienes production from human basophils derived from atopic dermatitis donors. *Int Arch Allergy Immunol* 107: 587-591.
- Shimizu T et al., 1994. Leukotriene B₄ and C₄ generation by blood leukocytes after *ex vivo* stimulation by Ca-ionophore and opsonized zymosan in children with atopic dermatitis. *Pediatr Allergy Immunol* 5: 95-99.
- Spergel JM and Paller ASS. 2003. Atopic dermatitis and the atopic march. *J Allergy Clin Immunol*. 112:S118-S127.
- Steinke JW and Boorish L. 2004. Leukotriene receptors in rhinitis and sinusitis. *Curr Allergy Asthma Rep* 4: 217-223.
- Stelmach I et al. 2002. A randomized, doubleblind trial of the effect of treatment with montelukast on bronchial hyperresponsiveness and serum eosinophilic cationic protein (ECP), soluble interleukin 2 receptor (sIL-2R), IL-4, and soluble intercellular adhesion molecule 1 (sICAM-1) in children with asthma. *J Allergy Clin Immunol* 109:257.
- Stulnig T.M. 2003. Immunomodulation by Polyunsaturated Fatty Acids: Mechanisms and Effects. *Int Arch Allergy Immunol* 132:310-321.
- Surette, M.E. et al. 2003a. Inhibition of Leukotriene Synthesis, Pharmacokinetics, and Tolerability of a Novel Dietary Fatty Acid Formulation in Healthy Adult Subjects. *Clinical Therapeutics* 25(3):948-971.

Surette, M.E. et al., 2003b. Inhibition of Leukotriene Synthesis by a Novel Dietary Fatty Acid Formulation in patients with Atopic asthma: A randomized, placebo-controlled, parallel-group, prospective trial. *Clinical Therapeutics* 25(3):972-979

Talbot SF et al. 1985. Accumulation of leukotriene C₄ and histamine in human allergic skin reactions. *J Clin Invest.* 76:650.

van Adelsberg J et al. 2003. Randomized controlled trial evaluating the clinical benefit of montelukast for treating spring seasonal allergic rhinitis. *Ann Allergy Asthma Immunol* 90:214-222.

Van Gool, CJAW et al., 2004. Oral essential fatty acid supplementation in atopic dermatitis – a meta-analysis of placebo-controlled trials. *Br J Dermatol* 150: 728-740.

Veien, NK et al. 2005. Montelukast treatment of moderate to severe atopic dermatitis in adults: A randomized, double-blind, placebo-controlled trial. *J Am Acad Dermatol* 53: 147-149.

Vladich FD et al., 2005. IL-13 R1130Q, a common variant associated with allergy and asthma, enhances effector mechanisms essential for human allergic inflammation. *J Clin Invest* 115: 747-754.

Wenzel SE. 1988. New approaches to anti-inflammatory therapy for asthma. *Am J Med* 104: 287-300.

Wenzel SE. 2003. The role of leukotrienes in asthma. *Prost. Leuk Essent Fatty Acids* 69:145-155.

Westbroek J. and Pasma HR. 2000. Effects of 2 weeks of treatment with fluticasone propionate 100 mcg b.d. by comparison with zafirlukast 20 mg b.d. on bronchial hyper-responsiveness in patients with mild to moderate asthma. *Respir Med* 94:112-118.

Woodmansee DP and Simon RA. 1999. A pilot study examining the role of zileuton in atopic dermatitis. *Ann Allergy Asthma Immunol* 83:548-552.

Yanase DJ and David-Bajar K. 2001. The leukotriene antagonist montelukast as a therapeutic agent for atopic dermatitis. *J Am Acad Dermatol* 44: 89-93.

Yoshida S et al. 2000. Effect of pranlukast on bronchial inflammation in patients with asthma. *Clin Exp Allergy* 30:1008-1014.

Zabawski EJ, Jr. et al., 1999. Treatment of atopic dermatitis with zafirlukast. *Correspondence to Dermatology Online Journal* 5: 10.

Zanconato S et al. 2004. Leukotrienes and 8-isoprostane in exhaled breath condensate of children with stable and unstable asthma. *J Allergy Clin Immunol.* 113:257-63.

Ziboh VA. 1996. The biological/nutritional significance of γ -linolenic acid in the epidermis: metabolism and generation of potent biological modulators. Chapter 10 in Huang, YS and Mills DE (Eds). *Gamma Linolenic Acid*. AOCS Press. Champaign, IL.

Ziboh VA and Chapkin 1987. *Arch. Derm* 123:1686-1690.

Ziboh VA and Fletcher MP. 1992. Dose-response effects of dietary gamma-linolenic acid-enriched oils on human polymorphonuclear-neutrophil biosynthesis of leukotriene B₄. *Am J Clin Nutr* 55:39-45.

Ziboh VA, Miller CC and Cho Y. 2000. Metabolism of polyunsaturated fatty acids by skin epidermal enzymes: generation of anti-inflammatory and antiproliferative metabolites. *Am J Clin Nutr* 71(suppl): 361A-366S.

Ziboh et al., 2004. Suppression of leukotriene B₄ generation by ex-vivo neutrophils isolated from asthma patients on dietary supplementation with gamma-linolenic acid-containing borage oil: possible implication in asthma. *Clin Devel Immunol* 11: 13-21.

Zsuzsanna C. et al. 2002. Increased Leukotrienes in Exhaled Breath Condensate in Childhood Asthma. *Am J Respir Crit Care Med* 166:1345-1349.